

## Albumin binding as a potential biomarker of exposure to moderately low levels of organophosphorus pesticides

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### Abstract

We have evaluated the potential of plasma albumin to provide a sensitive biomarker of exposure to commonly used organophosphorus pesticides in order to complement the widely used measure of acetylcholinesterase (AChE) inhibition. Rat or human plasma albumin binding by tritiated-diisopropylfluorophosphate ( $^3\text{H}$ -DFP) was quantified by retention of albumin on glass microfibre filters. Preincubation with unlabelled pesticide *in vitro* or dosing of F344 rats with pesticide *in vivo* resulted in a reduction in subsequent albumin radiolabelling with  $^3\text{H}$ -DFP, the decrease in which was used to quantify pesticide binding. At pesticide exposures producing approximately 30% inhibition of AChE, rat plasma albumin binding *in vitro* by azamethiphos (oxon), chlorfenvinphos (oxon), chlorpyrifos-oxon, diazinon-oxon and malaoxon was reduced from controls by  $9 \pm 1\%$ ,  $67 \pm 2\%$ ,  $56 \pm 2\%$ ,  $54 \pm 2\%$  and  $8 \pm 1\%$ , respectively. After 1 h of incubation with  $19 \mu\text{M}$   $^3\text{H}$ -DFP alone, the level of binding to rat or human plasma albumins reached 0.011 or 0.039 moles of DFP per mole of albumin, respectively. This level of binding could be further increased by raising the concentration of  $^3\text{H}$ -DFP, increasing the  $^3\text{H}$ -DFP incubation time, or by substitution of commercial albumins for native albumin. Pesticide binding to albumin was presumed covalent since it survived 24 h dialysis. After dosing rats with pirimiphos-methyl (dimethoxy) or chlorfenvinphos (oxon) (diethoxy) pesticides, the resultant albumin binding were still significant 7 days after dosing. As *in vitro*, dosing of rats with malathion did not result in significant albumin binding *in vivo*. Our results suggest albumin may be a useful additional biomonitor for moderately low-level exposures to several widely used pesticides, and that this binding differs markedly between pesticides.

**Keywords:** Albumin, organophosphorus pesticide, chlorfenvinphos (oxon), pirimiphos-methyl, chlorpyrifos-oxon, diazinon-oxon

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### Introduction

Exposure to organophosphorus (OP) pesticides can be estimated indirectly by inhibition of erythrocyte acetyl- or plasma butyryl-cholinesterase, or by measurement of urinary metabolites. In addition, a number of immune assays have been developed

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that are sufficiently sensitive to measure OP pesticides directly (Zhang et al. 2007). However, none of these approaches are ideal. Most OP pesticides are rapidly metabolised and eliminated from the body post-exposure. Cholinesterase inhibition, although longer lasting, does not discriminate between different inhibitors. Similarly, OP pesticides with very different toxic potential can share the same urinary metabolites, and these are also relatively rapidly eliminated post-exposure. It can, however, be important to be able to discriminate between exposures to different specific OP pesticides, because their action on targets such as neuropathy target esterase (Lotti 2002, Lotti & Moretto 2005), acylpeptide hydrolase (Richards et al. 2000) or other serine hydrolases (Ray & Richards 2001, Casida & Quistad 2004) does not parallel their actions on acetylcholinesterase (AChE). This diversity of action limits the value of AChE inhibition when used as the sole index of exposure. In addition, there are other forms of ill-health that have been associated with low-level exposure to OPs for which a molecular target has not yet been identified (Pope 1999, Ray & Richards 2001, Lotti 2002, Abou-Donia 2003, Kamel et al. 2005, Costa 2006), and for which it cannot be assumed that there is a parallel structure-activity relationship to that seen for AChE inhibition. Hence there is value in developing other additional biomarkers that have the potential to identify specific OP exposures. For example, acylpeptide hydrolase has a similar sensitivity to diisopropylfluorophosphate (DFP) exposure *in vitro* as butyrylcholinesterase (BuChE), but a markedly lower rate of spontaneous reactivation *in vivo*, indicative of its potential as a monitor of previous DFP or OP exposures (Quistad et al. 2005). This diversity of biological targets has potential to be exploited by use of a spectrum of biomarkers to 'fingerprint' different OP exposures.

Another potential biomarker for discriminating OP pesticide exposures is serum albumin. Albumin is adducted by DFP, and has been shown to bind a large number of OP pesticides *in vitro* – at least when incubated with relatively high pesticide to albumin molar ratios well beyond those likely to be encountered *in vivo* (Peeples et al. 2005). However, binding of pesticides to albumin may not always be covalent, notably for phosphorothioates (Mourik & Jong 1978, Maliwal & Guthrie 1981, Sultatos et al. 1984). We have shown that albumin does display a sensitive and compound specific OP pesticide binding at relatively low-exposure levels producing no more than 30% inhibition of AChE activity (Carter et al. 2007) – a level commonly considered to represent a just sub-symptomatic exposure for pesticide operators (Gallo & Lawryk 1991). An additional aspect of the binding of albumin by OPs is that albumin could act as a significant decoy target, reducing toxicity by limiting the access of OPs to functional targets in brain and muscle (Qiao et al. 2001).

Human serum albumin is the major blood protein, constituting 47% and 56% of rat and human plasma protein, respectively (Davies & Morris 1993, Peters 1996). Albumin functions to maintain colloidal osmotic pressure and pH, and is able to bind and transport both endogenous compounds, such as fatty acids and bilirubin, and a wide variety of exogenous compounds and drugs (Peters 1996). With its high concentration in blood, and a half-life of approximately 20 days in humans and 3 days in rats (Katz et al. 1961, Peters 1996), albumin displays suitable traits for a biomarker. However, little is known about the potential for albumin binding by pesticides at toxicologically relevant exposures, or of the susceptibility of OP-bound albumin to elimination and spontaneous reactivation *in vivo*. We have addressed these issues by evaluating the binding of rat and human plasma albumins *in vitro* by commonly used

OP pesticides when present as their biologically active (oxon) forms: azamethiphos (oxon), chlorfenvinphos (oxon), chlorpyrifos-oxon, diazinon-oxon and malaoxon. In addition, we have dosed rats with chlorfenvinphos (oxon), and the thion forms of two pesticides – pirimiphos-methyl and malathion – for bioactivation *in vivo* to their active oxon counterparts. Our results demonstrate differential pesticide binding of albumin both *in vitro* and *in vivo* at relatively low-level exposures. We further establish that rat plasma albumin binding by bioactivated pirimiphos-methyl (dimethoxy-) or chlorfenvinphos (oxon) (diethoxy-) *in vivo* results in OP-albumin binding that is still significant 7 days post-dosing, with a half-life similar to that for native rat albumin in either case. Our demonstration of significant albumin binding at sub-symptomatic exposure levels suggests that OP-albumin binding may indeed be suitable for exploitation as a reasonably long-lived biomarker of exposure to several OPs.

## Materials and methods

The organophosphorus pesticides azamethiphos (oxon) (*S*-6-chloro-2,3-dihydro-2-oxo-1,3-oxazolo[4,5-*b*]pyridin-3-ylmethyl *O,O*-dimethyl phosphorothioate), chlorfenvinphos (oxon) (2-chloro-1-(2,4-dichlorophenyl)vinyl diethyl phosphate) and malathion (diethyl (dimethoxyphosphinothioylthio)succinate) and its corresponding oxon (malaoxon) were bought from QMX Laboratories Ltd., Thaxted, UK. All compounds were at 95–99.5% purity. Chlorpyrifos (*O,O*-diethyl-*O*-(3,5,6-trichloro-2-pyridyl)phosphorothioate) and diazinon (*O,O*-diethyl *O*-2-isopropyl-6-methylpyrimidin-4-yl phosphorothioate) as their corresponding oxons, and pirimiphos-methyl (*O*-2-diethylamino-6-methylpyrimidin-4-yl *O,O*-dimethyl phosphorothioate) were purchased from Greyhound Laboratories, Birkenhead, UK. All compounds were at 97.2–99.4% purity. For *in vitro* assays, pesticides were prepared as 100 mM stock solutions in ethanol (Sigma, HPLC grade, <0.10% water), except azamethiphos (oxon) which was at 50 mM, and were stored at 4°C for up to 2 weeks. Pesticides were diluted in phosphate-buffered saline to required concentrations just prior to use. Tritiated-diisopropylfluorophosphate (<sup>3</sup>H-DFP) at a specific activity of 150 GBq mmol<sup>-1</sup> was purchased from Perkin Elmer, Boston, USA. The commercially purified albumins used for *in vitro* assays; rat albumin (A6272, purity >96%) and human albumin (A1653, purity 96–99%) were purchased from Sigma as were dithiothreitol (DTT), polyethylenimine solution and recombinant human AChE (C1682).

## Blood and tissue preparations

Fifty-two male F344 strain rats weighing between 200 and 230 g were used for experiments. Rats were maintained in cages (four per cage) under controlled temperature (21 ± 1°C) and light (16 h light/8 h dark cycle) with *ad libitum* access to food intake and water. All animal procedures were approved by the University of Nottingham Local Ethical Review Committee and were carried out in accordance with the Animals Scientific Procedures Act (UK) 1986.

Rats were dosed orally by gavage with OPs in arachis oil at 25% of their respective LD<sub>50</sub> values. Thus 15 rats were given 354 mg kg<sup>-1</sup> pirimiphos-methyl; 10 rats 2.5 mg kg<sup>-1</sup> chlorfenvinphos (oxon); and 5 rats 450 mg kg<sup>-1</sup> malathion. Control animals were given 1 ml kg<sup>-1</sup> arachis oil only (12 rats). In an additional experiment to investigate different OP pesticide doses, 5 rats were given 1.25 mg kg<sup>-1</sup> chlorfenvinphos (oxon)

(12.5% of the LD<sub>50</sub>), and 5 others 8.0 mg kg<sup>-1</sup> chlorfenvinphos (oxon) (80% of the LD<sub>50</sub>). One day after dosing rats were anaesthetised with isoflurane and 100–500 µl of blood removed from a tail vein. Alternatively, 1, 3 or 7 days after rat dosing approximately 5 ml of blood was removed by terminal intracardiac puncture. Heparinised blood was retained on ice before centrifugation at 2000g for 10 min at 4°C to pellet the red blood cells. The supernatant plasma was removed and stored at -80°C, and likewise the erythrocytes were decanted and stored at -80°C until required. Control human blood (taken with University of Nottingham Ethical Review Committee approval) from one of the authors (male, 39 years of age) was collected into heparin and similarly centrifuged to prepare erythrocytes and plasma.

Thymus and brain tissues were removed from control or dosed rats after saline perfusion, as described in Carter et al. (2007).

#### *Protein concentrations*

Protein concentrations were measured using the DC Protein assay (Biorad) using bovine serum albumin as a protein standard.

#### *Acetylcholinesterase measurements*

Inhibition of erythrocyte, thymus, or brain tissue AChE activity was measured based upon the spectrophotometric method described by Ellman et al. (1961). Spectrophotometric measurements were conducted at 412 nm in a Perkin Elmer Lambda 2S spectrophotometer operated using UV KinLab software. Replicate analyses for each assay sample point were conducted for 5 min at 37°C, with the rate of production of 5-thio-2-nitro-benzoic acid calculated, and averaged for the replicate readings. Erythrocytes were diluted 1:100 in 10 mM Tris/HCl pH 8.0 buffer for AChE measurements, brain was diluted 1:10 with the same buffer, and thymus tissue was used without dilution.

#### *Vacuum filtration of radiolabelled albumin and quantitation of radiolabelling*

Plasma radiolabelled with <sup>3</sup>H-DFP was separated by filtration on GF/B (25 mm) glass microfibre filters (Whatman) using a vacuum manifold. Filters were initially charged by soaking in 0.3% (w/v) polyethylenimine for 30 min. Charged filters were then applied to a vacuum manifold and washed with 10 mM Tris/HCl pH 8.0. Typically 10 µg of plasma proteins was loaded onto a single filter, washed with 50 ml of 10 mM Tris/HCl pH 8.0, air dried, and then counted for radioactivity within inserts containing 3 ml of scintillant. All assay points were performed in duplicate. Tritiated-DFP which was retained on the membranes in the absence of plasma proteins; approximately 1000 dpm, constituting 15 ± 0.3% of that for control plasma radiolabelling, was used as a blank and subtracted from all values.

#### *One-dimensional SDS-PAGE and albumin autoradiography*

Typically 20 µg of plasma was incubated with pesticide or phosphate-buffered saline as solvent for 20 min at room temperature. Proteins were then radiolabelled by incubation in a final volume of 130 µl with 19 µM <sup>3</sup>H-DFP (final concentration) for 1 h at 37°C typically at a molar ratio of <sup>3</sup>H-DFP:albumin of 8:1, or as described in

individual figure legends. Proteins were heated for 10 min at 70°C in sample buffer (Novex) containing 100 mM DTT. Proteins were separated on 4–12% Bis-Tris (Novex) gels run with 3-(N-morpholino)propanesulfonic acid (MOPS) buffer, then electroblotted onto a polyvinylidene difluoride membrane, cross-linked, and  $^{14}\text{C}$  markers applied to the blot to identify the positions of the molecular weight standards. Blots were subjected to 7–24 h of autoradiography within a highly sensitive microchannel plate detector as detailed in Carter et al. (2007). Protein radiolabelled bands from autoradiographic images were quantified using Quant scan software (Beta autoradiographic image acquisition software), with band intensities (in pixels) plotted using excel to determine relative radiolabel incorporations.

### Statistical analysis

For comparison of the levels of radioactivity incorporated into albumin, a one-way analysis of variance (ANOVA) with *post-hoc* test was performed (Bonferroni's multiple comparison test).

## Results

### *Albumin is differentially bound by organophosphorus pesticide-oxons in vitro at toxicologically relevant doses*

Rat plasma was diluted to 320  $\mu\text{g ml}^{-1}$  and preincubated with pesticide-oxon or solvent for 20 min *in vitro* at concentration/times confirmed to produce approximately 30% inhibition of thymus tissue AChE activity (Carter et al. 2007). The actual concentrations used were: azamethiphos (oxon) 0.12  $\mu\text{M}$ , chlorfenvinphos (oxon) 1.5  $\mu\text{M}$ , chlorpyrifos-oxon 0.03  $\mu\text{M}$ , diazinon-oxon 0.04  $\mu\text{M}$  and malaoxon 0.1  $\mu\text{M}$ . Residual active hydrolase groups were then adducted by incubation with 19  $\mu\text{M}$   $^3\text{H}$ -DFP for 1 h at 37°C. Radiolabelled proteins (typically 10  $\mu\text{g}$  per data point) were loaded onto glass microfibre filters and washed extensively to remove extraneous unincorporated  $^3\text{H}$ -DFP. After drying, filters were counted for radioactivity and both the maximal level of  $^3\text{H}$ -DFP incorporation and the level of pesticide binding (fall in radioactivity) quantified (Figure 1A). Alternatively, radiolabelled plasma proteins were subjected to one-dimensional SDS-PAGE and autoradiography. After radiolabelling of 20  $\mu\text{g}$  of rat plasma only a single radiolabelled protein was present after protein separation by denaturing SDS-PAGE. This radiolabelled albumin migrated as a discrete band of approximate molecular weight 66 kDa at the leading edge of the main Coomassie-stainable albumin (Figure 1B). This band was confirmed as serum albumin by its removal by immunoprecipitation; localisation by Western blotting using an antialbumin antibody (Autogen bioclear, ABN 192); and also by peptide mass fingerprinting of the excised protein by matrix-assisted laser-desorption time-of-flight mass spectrometry (results not included).

From either quantitation of  $^3\text{H}$ -DFP radiolabelled albumin retained on microfibre filters (Figure 1A), or from autoradiography of gel-resolved protein (Figure 1B), plasma albumin was shown to be bound *in vitro* by the oxons of chlorfenvinphos ( $67 \pm 2\%$ ), chlorpyrifos ( $56 \pm 2\%$ ) and diazinon ( $54 \pm 2\%$ ), but not azamethiphos ( $9 \pm 1\%$ ) or malathion ( $8 \pm 1\%$ ) relative to controls. Comparison of the proportion of albumin radiolabelled (by autoradiography) with the total protein present (visualised by Coomassie blue protein staining) showed that only a small proportion of albumin

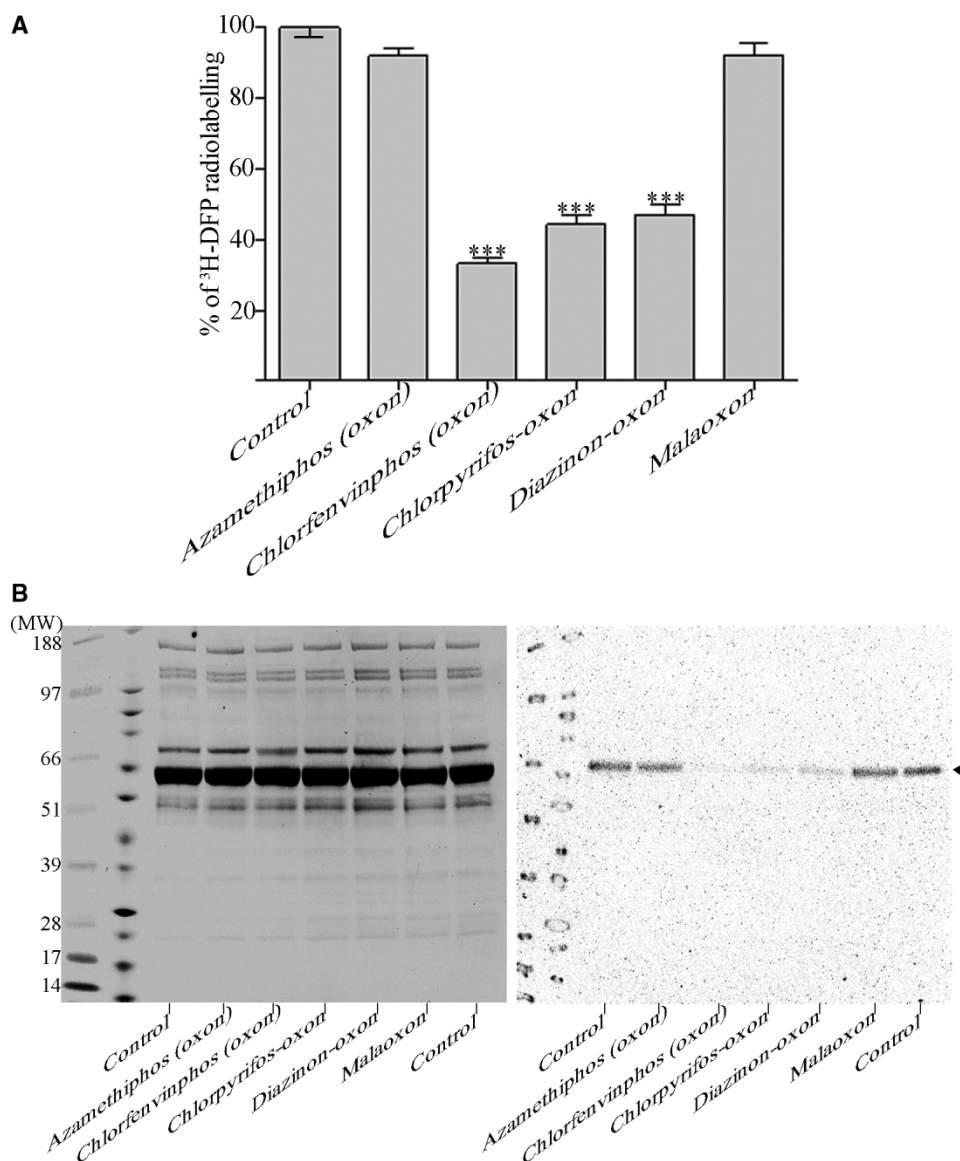


Figure 1.  $^3\text{H}$ -DFP incorporation into rat plasma albumin after preincubation with pesticide-oxons. Rat plasma was incubated with pesticide-oxons or solvent at 20 min thymus tissue AChE  $\text{IC}_{50}$  concentrations. Reactive hydrolase groups were then adducted by incubation with  $^3\text{H}$ -DFP for 1 h at  $37^\circ\text{C}$ . (A) Ten micrograms of radiolabelled plasma was loaded onto glass microfibre filters, washed, and the incorporation of  $^3\text{H}$ -DFP determined. Results are presented as the mean  $\pm$  standard error from at least 16 independent experiments with each pesticide. Results were significantly different from controls with chlorfenvinphos (oxon), chlorpyrifos-oxon, and diazinon-oxon ( $***p < 0.001$ ). (B) Twenty micrograms of radiolabelled plasma was resolved by SDS-PAGE, and then transferred to a polyvinylidene difluoride membrane. Proteins were stained with Coomassie brilliant blue (left panel) or autoradiographed (right panel). The positions of protein molecular weight markers are shown in the first two lanes of the gel and blot. A single radiolabelled protein band was evident (marked with an arrowhead) which superimposed with the leading edge of the albumin protein band detected from protein staining. Incubation with the pesticide-oxons of chlorfenvinphos, chlorpyrifos, and diazinon significantly reduced  $^3\text{H}$ -DFP incorporation into this radiolabelled protein.



molecules were bound by  $^3\text{H}$ -DFP under the conditions employed. The level of  $^3\text{H}$ -DFP binding was quantified as approximately 1% of albumin molecules using the microfibre binding and quantitation method. This calculation assumes that protein recovery by this method was 100%, and so may have somewhat underestimated the true stoichiometry, but these filters are known to achieve high protein recovery (Bruns et al. 1983).

To validate this stoichiometry calculation by an independent means, we also radiolabelled rat plasma albumin with  $^3\text{H}$ -DFP alongside  $^3\text{H}$ -DFP radiolabelled recombinant AChE. Radiolabelled proteins were then resolved by gel electrophoresis and subjected to autoradiography using a microchannel plate detector, a device with known autoradiographic signal linearity over six orders of magnitude (Richards & Lees 2002). Both proteins have a similar molecular weight but AChE can incorporate organophosphates stoichiometrically, i.e. one mole of organophosphate bound per mole of AChE (Raveh et al. 1989). A signal of 3640 arbitrary pixel intensity units was generated from 24 h autoradiography of 0.2  $\mu\text{g}$  of  $^3\text{H}$ -DFP radiolabelled AChE, whereas 3000 intensity units arose from 20  $\mu\text{g}$  of  $^3\text{H}$ -DFP radiolabelled albumin (images not included). Assuming 100% binding of  $^3\text{H}$ -DFP to AChE,  $^3\text{H}$ -DFP binding to albumin was therefore quantified as 0.82% of available albumin molecules, a value close to the 1% binding evidenced from retention of  $^3\text{H}$ -DFP radiolabelled albumin on microfibre filters.

Since  $^3\text{H}$ -DFP binding to albumin survived denaturing SDS-PAGE, it was presumed covalent in nature and not transient or readily reversible. That this was also true of the unlabelled test pesticides was evidenced by the reduction in  $^3\text{H}$ -DFP labelling of albumin produced by preincubation with pesticide being unchanged after extensive (24 h) dialysis to remove free OP prior to  $^3\text{H}$ -DFP radiolabelling (results not included).

*The absolute level of in vitro albumin binding by  $^3\text{H}$ -DFP was related to the  $^3\text{H}$ -DFP:albumin molar ratio, time of incubation, and albumin source and purification*

We sought an explanation of this relatively low  $^3\text{H}$ -DFP albumin binding stoichiometry, since other literature reports have suggested higher reactivity potential (Murachi 1963, Means & Wu 1979). If the reaction with  $^3\text{H}$ -DFP was dependent upon both the molar ratio of  $^3\text{H}$ -DFP to albumin, and also the time of incubation with  $^3\text{H}$ -DFP, then increasing either should increase the radiolabelling of albumin. We incubated diluted rat plasma (320  $\mu\text{g ml}^{-1}$  final concentration) with  $^3\text{H}$ -DFP at a final concentration range of 5.5  $\mu\text{M}$  to 19  $\mu\text{M}$ . At this rat plasma dilution, albumin is at a concentration of approximately 2.3  $\mu\text{M}$  giving a molar ratio of approximately 2:1 to 8:1 moles of  $^3\text{H}$ -DFP:moles of albumin, respectively. Increasing the molar ratio of  $^3\text{H}$ -DFP to plasma albumin did indeed increase the level of  $^3\text{H}$ -DFP binding (Table I) (control row). Across this range of  $^3\text{H}$ -DFP:albumin molar ratios, preincubation with pesticides *in vitro* (under conditions producing approximately 30% thymus tissue AChE inhibition) produced a similar differential binding pattern as that seen with 19  $\mu\text{M}$   $^3\text{H}$ -DFP (Table I, and Figure 1A, B). The absolute magnitude of the pesticide binding increased with  $^3\text{H}$ -DFP concentration, although not proportionately. This suggested that not all of the additional pesticide binding sites disclosed by the higher  $^3\text{H}$ -DFP concentrations were targeted by the pesticides (see Table I).

Table I. Stiochiometry of  $^3\text{H}$ -DFP incorporation into rat plasma albumin. Rat plasma albumin ( $2.3\ \mu\text{M}$ ) was incubated with solvent or pesticide-oxons at 20 min AChE  $\text{IC}_{30}$  concentrations, and then radiolabelled with 5.5, 10 or  $19\ \mu\text{M}$   $^3\text{H}$ -DFP. Both the increase in albumin radiolabelling from increasing  $^3\text{H}$ -DFP concentration, and the level of pesticide binding relative to controls was determined by quantifying the level of radiolabelled albumin retained on glass microfibre filters.

Preincubation	Stoichiometry of subsequent $^3\text{H}$ -DFP incorporation (% moles $^3\text{H}$ -DFP:moles albumin) Concentration of $^3\text{H}$ -DFP employed		
	5.5 $\mu\text{M}$ ( $n=4$ )	10 $\mu\text{M}$ ( $n=5$ )	19 $\mu\text{M}$ ( $n=16$ )
Solvent (control)	$0.55 \pm 0.01$ (100%)	$0.70 \pm 0.01$ (100%)	$1.08 \pm 0.03$ (100%)
Azamethiphos (oxon)	$0.47 \pm 0.03$ ( $85 \pm 1.1\%$ )	ND	$0.99 \pm 0.03$ ( $91 \pm 1.1\%$ )
Chlorfenvinphos (oxon)	$0.09 \pm 0.01$ ( $16 \pm 3.7\%$ )	$0.16 \pm 0.01$ ( $22 \pm 2.8\%$ )	$0.35 \pm 0.02$ ( $33 \pm 2.0\%$ )
Chlorpyrifos-oxon	$0.12 \pm 0.01$ ( $22 \pm 1.6\%$ )	$0.25 \pm 0.03$ ( $35 \pm 2.0\%$ )	$0.47 \pm 0.03$ ( $44 \pm 1.7\%$ )
Diazinon-oxon	$0.12 \pm 0.01$ ( $22 \pm 1.6\%$ )	$0.28 \pm 0.02$ ( $40 \pm 1.8\%$ )	$0.49 \pm 0.03$ ( $46 \pm 1.6\%$ )
Malaoxon	$0.53 \pm 0.01$ ( $96 \pm 1.0\%$ )	ND	$0.99 \pm 0.03$ ( $92 \pm 1.1\%$ )

ND, not determined.

To examine the effect of increasing the time of incubation of  $^3\text{H}$ -DFP with plasma albumin at a given  $^3\text{H}$ -DFP:albumin molar ratio (8:1),  $^3\text{H}$ -DFP was incubated with plasma albumin at  $37^\circ\text{C}$  for a time course of up to 4 days. Plasma samples were removed at time points during the incubation period, and the level of  $^3\text{H}$ -DFP incorporation into albumin quantified by counting  $^3\text{H}$ -DFP-radiolabelled albumin retained on microfibre filters. As  $^3\text{H}$ -DFP is unstable after 24 h in aqueous solutions (Hobbiger 1951) additional  $^3\text{H}$ -DFP was added to the reaction mixture at intervals of 24 h. The level of  $^3\text{H}$ -DFP incorporation into rat plasma albumin increased in a roughly linear fashion over the first 4 h but without a further increase at 24 h, indicative of  $^3\text{H}$ -DFP hydrolysis. However, after supplementation with additional  $^3\text{H}$ -DFP after the 24 h time point, and daily thereafter, an approximately linear  $^3\text{H}$ -DFP incorporation was seen (Figure 2). This confirmed that albumin could be radiolabelled at a higher stiochiometry if incubation times with  $^3\text{H}$ -DFP were extended. This increase in  $^3\text{H}$ -DFP radiolabelling was not a result of denaturation of albumin over the time course, since preincubation of plasma albumin alone for 4 days prior to radiolabelling had no influence on the level of subsequent  $^3\text{H}$ -DFP incorporation, suggesting that the binding site was still intact and that the relatively low binding stiochiometry was indeed a result of slow  $^3\text{H}$ -DFP binding (results not included).

Other groups have analysed the binding of DFP and/or pesticides *in vitro* using commercially purified bovine or human albumin which are essentially fatty acid free (Means & Wu 1979, Hagag et al. 1983, Peeples et al. 2005, Li et al. 2007). Commercially available albumins (Fraction V) undergo purification conditions that include a series of alcohol fractionations (Peters 1996), conditions that would be expected to induce albumin conformational changes which may result in increased susceptibility of the active site tyrosine to  $^3\text{H}$ -DFP binding. To test this hypothesis we radiolabelled human plasma, and commercially purified rat and human albumins with  $19\ \mu\text{M}$   $^3\text{H}$ -DFP for 1 h at  $37^\circ\text{C}$ , and compared the level of radioactivity incorporated with that from our rat plasma studies (Table II). Interestingly, human plasma albumin was radiolabelled by  $^3\text{H}$ -DFP to a higher stiochiometry than rat plasma approaching 4% of albumin molecules, but the absolute reduction in labelling seen after OP preincubations was not significantly greater, suggesting that these additional  $^3\text{H}$ -DFP binding sites were not sensitive to the pesticides tested. For both rat and human



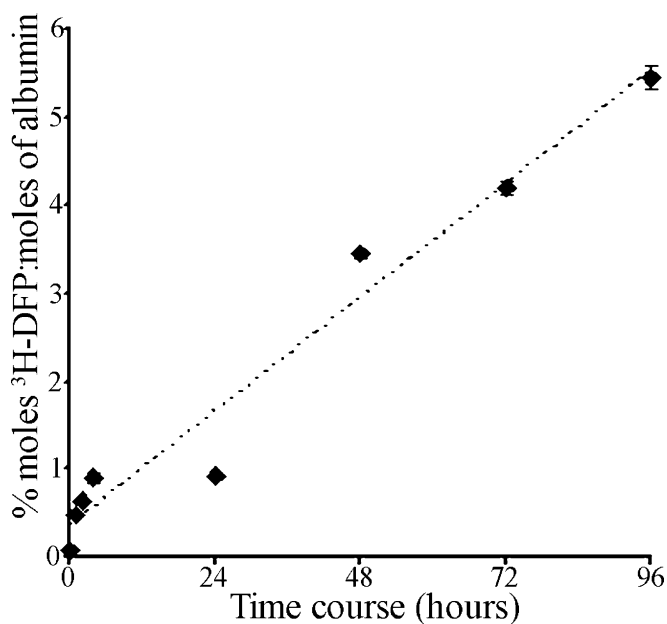


Figure 2. Time course of  $^3\text{H}$ -DFP incorporation into rat plasma albumin. Rat plasma (300  $\mu\text{g}$ ) was reacted at  $37^\circ\text{C}$  with 19  $\mu\text{M}$   $^3\text{H}$ -DFP (19 nmoles) in a buffer of 10 mM Tris/HCl pH 8.0 containing 1 mM EDTA, 5 mM DTT, 5% glycerol (1000  $\mu\text{l}$  final volume). At intervals of 1, 2, 4, 24, 48, 72, and 96 h, replicate 50  $\mu\text{l}$  samples were removed and the level of  $^3\text{H}$ -DFP incorporated into albumin quantified by counting the radiolabelled albumin retained on glass microfibre filters. To counter  $^3\text{H}$ -DFP hydrolysis during the time course, the reaction mixture was supplemented with 7.2 nmoles of  $^3\text{H}$ -DFP after 24 h, 4.8 nmoles of  $^3\text{H}$ -DFP after 48 h, and 2.4 nmoles of  $^3\text{H}$ -DFP after 72 h. Data points are mean  $\pm$  standard deviation from four independent experiments.

commercial albumins, the level of  $^3\text{H}$ -DFP binding stoichiometry increased by approximately 5-fold over the native forms, suggesting that commercial albumin purification may result in albumin molecules with increased accessibility of their active site to binding by  $^3\text{H}$ -DFP.

Table II. Stoichiometry of  $^3\text{H}$ -DFP incorporation into rat and human albumins. Rat or human albumin (1–2  $\mu\text{M}$ ) was incubated with solvent or pesticide-oxons at 20 min AChE  $\text{IC}_{50}$  concentrations, and then radiolabelled with 19  $\mu\text{M}$   $^3\text{H}$ -DFP. The level of albumin radiolabelling was quantified by counting radiolabelled albumin retained on glass microfibre filters. For rat plasma albumin,  $n = 16$ , and for all other data points  $n = 4$ –6 experiments.

Agent	Stoichiometry of $^3\text{H}$ -DFP incorporation (% moles $^3\text{H}$ -DFP:moles albumin)			
	Rat plasma albumin	Human plasma albumin	Purified Rat albumin	Purified human albumin
Solvent (control)	$1.08 \pm 0.03$	$3.87 \pm 0.03$	$6.20 \pm 0.14$	$19.03 \pm 0.17$
Azamethiphos (oxon)	$0.99 \pm 0.03$	$3.67 \pm 0.06$	$5.94 \pm 0.30$	$18.65 \pm 0.26$
Chlorfenvinphos (oxon)	$0.35 \pm 0.02$	$2.96 \pm 0.07$	$4.83 \pm 0.11$	$16.50 \pm 0.09$
Chlorpyrifos-oxon	$0.47 \pm 0.03$	$2.97 \pm 0.06$	$5.20 \pm 0.10$	$17.32 \pm 0.33$
Diazinon-oxon	$0.49 \pm 0.03$	$3.24 \pm 0.17$	$5.32 \pm 0.18$	$17.63 \pm 0.31$
Malaoxon	$0.99 \pm 0.03$	$3.35 \pm 0.06$	$5.56 \pm 0.07$	$18.25 \pm 0.38$

*Albumin binding by pesticide-oxons in vitro was proportional to pesticide concentration, and was of a comparable sensitivity to pesticide binding of AChE*

The effect of varying concentrations of the albumin-binding pesticides chlorfenvinphos (oxon), chlorpyrifos-oxon and diazinon-oxon on the  $^3\text{H}$ -DFP radiolabelling of rat plasma albumin and the inhibition of AChE activity were examined in parallel incubations. Rat plasma (at a protein concentration of  $203\text{ }\mu\text{g ml}^{-1}$ ), or recombinant AChE, were preincubated with pesticides for 20 min over a concentration range of  $1\text{ nM} - 200\text{ }\mu\text{M}$  for chlorfenvinphos (oxon),  $1\text{ nM} - 100\text{ }\mu\text{M}$  for chlorpyrifos-oxon, and  $1\text{ nM} - 100\text{ }\mu\text{M}$  for diazinon-oxon. The level of AChE inhibition was quantified by the Ellman assay (Ellman et al. 1961). After pesticide preincubations rat plasma proteins were radiolabelled with  $17\text{ }\mu\text{M}$   $^3\text{H}$ -DFP for 1 h at  $37^\circ\text{C}$ , and the incorporation of radioactivity into albumin quantified by counting  $^3\text{H}$ -DFP-radiolabelled albumin retained on microfibre filters, and also visualised after autoradiography of SDS-PAGE separated proteins (Figure 3A–C).

Both albumin binding and AChE inhibition were proportional to pesticide concentrations. For chlorfenvinphos (oxon), the  $\text{IC}_{50}$  for AChE and albumin binding were approximately  $1.5$  and  $0.4\text{ }\mu\text{M}$ ; for chlorpyrifos-oxon,  $0.04$  and  $0.1\text{ }\mu\text{M}$ ; and for diazinon-oxon,  $0.5$  and  $2\text{ }\mu\text{M}$ , respectively. Thus albumin exhibited an *in vitro* sensitivity to binding by these pesticide-oxons that was similar to (and for chlorfenvinphos (oxon) greater than) that of AChE inhibition.

*Albumin is bound by pirimiphos-methyl and chlorfenvinphos (oxon) in vivo with half-lives comparable to that of native rat albumin*

Having established that albumin binding by certain pesticides is seen *in vitro* at toxicologically relevant pesticide exposures, we investigated albumin binding *in vivo*, and also whether pesticide binding to albumin is sufficiently stable to provide a biomonitor of pesticide exposure.

Rats were dosed with solvent (controls), or with pesticides at 25% of their  $\text{LD}_{50}$ :  $354\text{ mg kg}^{-1}$  pirimiphos-methyl (a thion for bioactivation to its oxon counterpart *in vivo*), and  $2.5\text{ mg kg}^{-1}$  chlorfenvinphos (oxon). Rats remained asymptomatic at these dose levels. Blood was removed at 1, 3, or 7 days after dosing, and plasma albumin radiolabelled with  $19\text{ }\mu\text{M}$   $^3\text{H}$ -DFP for 1 h at  $37^\circ\text{C}$ . The incorporation of radioactivity into albumin was quantified by counting  $^3\text{H}$ -DFP-radiolabelled albumin retained on microfibre filters (Figure 4A, B). In addition, radiolabelled albumin was resolved by SDS-PAGE and visualised by autoradiography. An example of the level of radiolabel incorporated into albumin in one rat from each group is also included in Figure 4A and B (lower panels). Erythrocytes from the rats were used to determine the corresponding level of AChE inhibition arising from each dose by the Ellman assay.

Figure 3. Comparison of pesticide binding of albumin with inhibition of AChE. Rat plasma ( $203\text{ }\mu\text{g ml}^{-1}$ ) was incubated with pesticide or solvent for 20 min at room temperature over the pesticide concentration ranges shown. Proteins were then radiolabelled with  $17\text{ }\mu\text{M}$   $^3\text{H}$ -DFP for 1 h at  $37^\circ\text{C}$ . The radiolabel incorporated into albumin was quantified by counting radiolabelled albumin retained on glass microfibre filters. The corresponding inhibition of AChE at each concentration was quantified using the Ellman assay, with the results presented graphically. Radiolabelling of albumin was also visualised after SDS-PAGE and autoradiography and is shown in the lower panels. (A) Chlorfenvinphos (oxon), (B) chlorpyrifos-oxon, (C) diazinon-oxon.

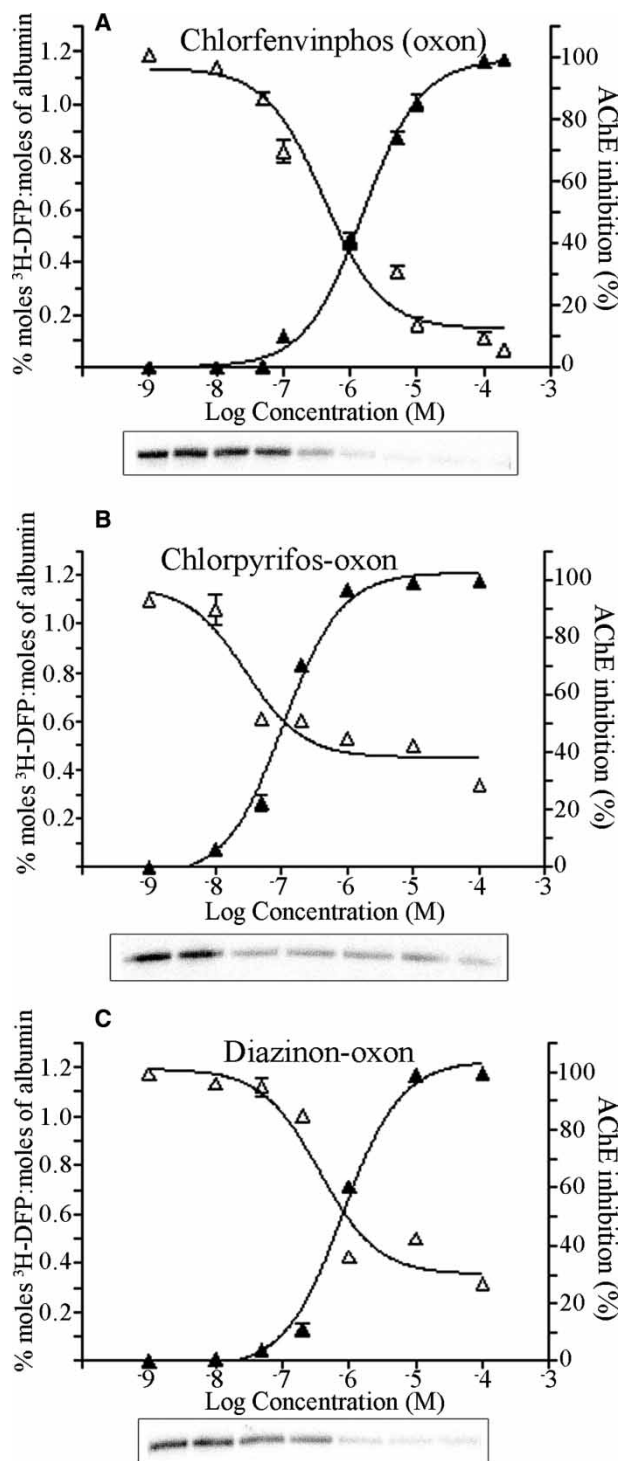


Figure 3 (Continued)

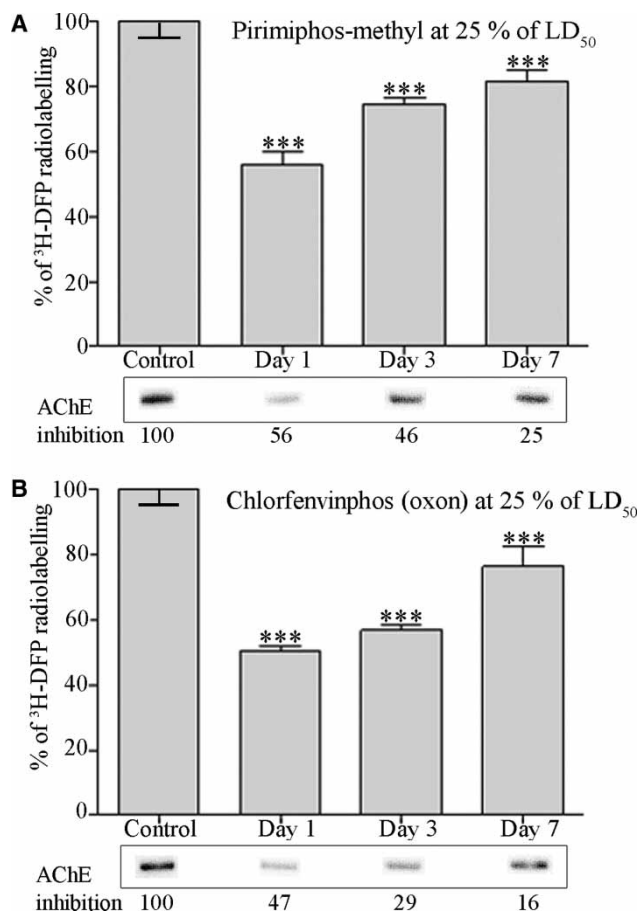


Figure 4. Quantitation of dimethoxy- and diethoxy-pesticide binding of albumin *in vivo*. Rats were treated with solvent (controls) or dosed with 25% of the LD<sub>50</sub> of pirimiphos-methyl (A), or 25% of the LD<sub>50</sub> of chlorfenvinphos (oxon) (B). After radiolabelling with <sup>3</sup>H-DFP the incorporation of radioactivity into albumin was quantified by counting the radiolabelled albumin retained on glass microfibre filters. Results are presented as the mean  $\pm$  standard error from at least four independent experiments from each rat. Results were significantly different from controls at 1, 3, and 7 days after dosing (\*\**p* < 0.001) for both pesticides. Radiolabelled albumin was also resolved by SDS-PAGE, and an example of the level of radioactivity incorporated into albumin for each dose condition displayed in the lower panels. The average level of erythrocyte AChE inhibition for each dosing condition is also included.

When plasma albumin was analysed 1 day after dosing, both pirimiphos-methyl and chlorfenvinphos (oxon) had inhibited approximately 50% of the erythrocyte AChE activity, with a corresponding inhibition of binding of approximately 50% of the <sup>3</sup>H-DFP-radiolabelled albumin molecules. This level was comparable to the 'maximal' pesticide binding achieved from our *in vitro* studies. Analysis of the elimination of pesticide binding to albumin over a 1 week post-dosing time course, revealed that both pirimiphos-methyl (dimethoxy-) and chlorfenvinphos (oxon) (diethoxy-) binding steadily returned toward control levels, but was still significant 3 and 7 days after dosing.

A natural logarithmic plot of the elimination of the pesticide binding to albumin for pirimiphos-methyl or chlorfenvinphos (oxon) indicated half-lives of at least 3 days (graph not included), suggesting that pesticide-bound albumin was not actively

degraded or unstable, but followed a similar rate of turnover to that described for native rat albumin (Peters 1996, Troester et al. 2002).

*Albumin is not bound by malathion in vivo but is maximally bound by chlorfenvinphos (oxon) even at low AChE inhibitions*

To establish whether the differential pesticide binding of albumin that we have documented *in vitro* was operable *in vivo*, rats were dosed with malathion (at approximately 25% of the LD<sub>50</sub>), a thion for bioactivation *in vivo* to its oxon counterpart, malaoxon – a pesticide that did not significantly bind albumin *in vitro*. In addition, to determine if albumin binding by chlorfenvinphos (oxon) was increased at a higher pesticide dose, or decreased with a lower dose, rats were given either 80% or 12.5% of the LD<sub>50</sub>. The higher dose, but not the lower dose, produced salivation and muscle fasciculation in all rats. Twenty-four hours after dosing rat blood was removed, and the level of albumin binding quantified by counting the <sup>3</sup>H-DFP-radiolabelled albumin retained on microfibre filters, and the corresponding levels of erythrocyte AChE inhibition determined by the Ellman assay (Figure 5). For each of the chlorfenvinphos (oxon) doses analysed, approximately 50% of the <sup>3</sup>H-DFP radiolabelled albumin molecules were bound, suggesting that this level of binding was also 'maximal' for the *in vivo* exposure duration. Moreover, even at the lowest AChE inhibition (22%) achieved with the lowest chlorfenvinphos (oxon) dose, albumin binding was still maximal, supporting our *in vitro* studies that chlorfenvinphos (oxon)

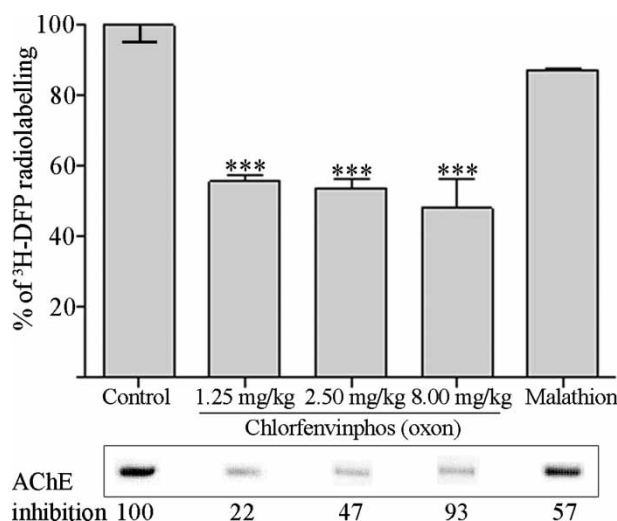


Figure 5. Albumin is maximally bound across a range of chlorfenvinphos (oxon) doses *in vivo*, but is not significantly bound by malathion *in vivo*. Rats were treated with solvent (controls) or dosed with 12.5%, 25%, and 80% of the LD<sub>50</sub> of chlorfenvinphos (oxon), or approximately 25% of the LD<sub>50</sub> of malathion. Twenty-four hours after dosing plasma albumin was radiolabelled with <sup>3</sup>H-DFP, and the incorporation of radioactivity into albumin quantified by counting the radiolabelled albumin retained on glass microfibre filters. Results are presented as the mean  $\pm$  standard error from at least four independent experiments from each rat. Results significantly different from controls are marked (\*\*\**p* < 0.001). Radiolabelled albumin was also visualised by autoradiography after SDS-PAGE, and an example of the level of radioactivity incorporated into albumin shown in the lower panels. The average level of erythrocyte AChE inhibition for each dosing condition is also included.

binding to albumin was at least, if not more, sensitive than binding to AChE. By contrast, no significant binding of albumin by malathion was evident (despite 57% AChE inhibition), also in agreement with our *in vitro* studies, that malathion, or its bioactivated oxon (malaoxon), were not capable of significantly binding albumin.

## Discussion

In order to assess the potential for albumin to act as a biomarker for a range of pesticides in common use within the UK or USA, we used DFP labelling to quantify the levels of pesticide binding to rat albumin primarily at sub-symptomatic OP exposure levels. After pesticide preincubations *in vitro* sufficient to produce 30% thymus tissue AChE inhibition, marked albumin binding was demonstrated with the oxons of chlorfenvinphos, chlorpyrifos and diazinon, but not with those of azamethiphos or malathion. This binding was confirmed as covalent by extensive dialysis prior to  $^3\text{H}$ -DFP incubation. The other major characterised  $^3\text{H}$ -DFP target proteins in plasma, AChE and BuChE, are at very low concentrations relative to albumin so, although they will also have been reacted with these pesticides, their contribution to the total level of binding calculated would be insignificant. For comparison, albumin is at an approximate plasma protein concentration of  $32\text{ mg ml}^{-1}$  in rats and  $42\text{ mg ml}^{-1}$  in humans ( $0.5\text{--}0.6\text{ mM}$ ) (Davies & Morris 1993), whereas the human concentration of plasma BuChE has been estimated at  $3.3\text{ }\mu\text{g ml}^{-1}$ , or approximately  $50\text{--}80\text{ nM}$  (Brimijoin & Hammond 1988, van der Schans et al. 2004). For AChE, the human plasma concentration has been estimated as  $8\text{ ng ml}^{-1}$  (Brimijoin & Hammond 1988). That the binding of pesticides to these two cholinesterases was not significant under our assay conditions was confirmed experimentally by preincubation of plasma with the AChE inhibitor eserine (Sigma E8375), or the BuChE inhibitor ethopropazine (Sigma L308765), which in neither case lowered radioactivity incorporated into plasma albumin from that of control levels (results not included).

In order to evaluate the functional significance of this albumin binding, it is necessary to estimate the absolute stoichiometry; however, as radiolabelled pesticides were not available, we achieved this indirectly via  $^3\text{H}$ -DFP binding. Clearly DFP is not an ideal ligand for this purpose, since after 1 h incubation with  $19\text{ }\mu\text{M}$   $^3\text{H}$ -DFP only approximately 1% of rat and 4% of human albumin molecules were radiolabelled via calculation of the retention of  $^3\text{H}$ -DFP-radiolabelled albumin on microfibre filters.

However, increasing the molar ratio of  $^3\text{H}$ -DFP to albumin, extending the time course with  $^3\text{H}$ -DFP or replacement of native with commercially purified albumin, all increased the level of  $^3\text{H}$ -DFP radiolabelling of albumin. This concurs with the results described by other groups detailing higher stoichiometries of binding after either prolonged and/or high concentrations of inhibitor used under assay conditions of above pH 8.0 which facilitate binding-site accessibility, and/or from incubations with commercially prepared albumins from which fatty acids have been stripped (Murachi 1963, Means & Wu 1979, Hagag et al. 1983, Li et al. 2007). However, this higher level of albumin binding would not be expected to be seen with native albumin *in vivo*.

The low stoichiometry of radiolabelling of albumin that we report may reflect an inaccessibility of the active site to adduction by  $^3\text{H}$ -DFP. In contrast to the more usual adduction of active-site serine hydrolases, DFP and OPs bind an active-site tyrosine in serum albumin (tyrosine 411 of human or rat albumin) (Sanger 1963, Means & Wu 1979, Black et al. 1999, Schopfer et al. 2005, Li et al. 2007). Since this active-site



tyrosine is also a point of ionic contact for associated molecules including both monounsaturated oleic acid and polyunsaturated arachidonic acid, two fatty acids that normally associate with albumin in serum (Petitpas et al. 2001), DFP binding may certainly be conformationally restricted by the presence of these or similar fatty acids (Means & Wu 1979, Petitpas et al. 2001), a restriction which may be lost in commercial albumin preparations. We did not investigate this further, concentrating our studies on interactions with the native, fatty acid bound albumin which is present *in vivo*.

Marked differences between the levels of binding of the different OPs to albumin were evident from both our *in vitro* and *in vivo* results, in contrast to the uniform binding shown by the majority of agents tested at the very high exposure levels used by others (Peeples et al. 2005). The structures of the OPs used in this study are shown in Figure 6, and divide into relatively low and high albumin-binding structures. Although these OPs were selected on the basis of pesticide usage rather than structure, it is apparent that those that possess an ester linkage to the leaving group readily bind albumin, whereas those with a thioester are virtually unreactive. Hence we might predict that other similarly structured dimethoxy triesters in which the leaving group is thioester linked, such as azinphos-methyl and possibly its diethoxy triester counterpart azinphos-ethyl, would be unreactive with albumin under conditions in which similarly structured esters were reactive. This prediction, like the mechanistic basis for albumin's differential pesticide reactivity, will need to be tested via further experimentation.

The functional significance of albumin binding by OPs is unclear at present. There are a number of enzymatic activities intrinsic to serum albumin that have the potential to counter certain xenobiotics, of which the OP-binding or fatty acid binding site, tyrosine 411, may also be an enzymatically active site residue (Means & Wu 1979, Hagag et al. 1983, Sultatos et al. 1984, Sogorb et al. 1998, Watanabe et al. 2000, Kragh-Hansen et al. 2002). However, the majority of these enzymatic activities are relatively weak, or only operate in the absence of fatty acids and/or at pHs above the normal physiological range (Ortigoza-Ferado et al. 1984). Nonetheless, irrespective of its relative enzymatic inabilities, albumin does constitute the major plasma protein; hence, if only 1% of albumin molecules in adults are physiologically active to sequester OPs or catalytically hydrolyse OPs (act as an A-esterase) or other xenobiotics, albumin will still exert considerable influence on the free circulatory OP/xenobiotic concentration, and therefore its tissue distribution, liberation, elimination, and thus detoxification. This has recently been substantiated, at least *in vitro*, by a demonstration of the detoxification of the carbamate pesticide carbaryl by bovine serum albumin at toxicologically relevant concentrations (Sogorb et al. 2007). It is noteworthy that this protection from neurotoxicity that binding of OPs to albumin affords, may well be compromised in the fetus and newborn since they possess lower concentrations of albumin and other serum proteins (Qiao et al. 2001).

The binding of OP pesticides to albumin will also have the potential to compete and/or displace endogenous and exogenous albumin-bound compounds, including an array of drugs that also bind albumin at the same region of the molecule, hence influencing drug metabolism and pharmacokinetics (Watanabe et al. 2000, Kragh-Hansen et al. 2002). Furthermore, since human albumin has the second highest number of allelic mutants after haemoglobin including residues adjacent to the OP-binding site (referenced at the ExPasy website: <http://au.expasy.org/uniprot/P02768>, or at <http://www.albumin.org/>), with some mutants influencing fatty acid (or drug) binding (Nielsen et al. 1997), then presumably the binding of OPs to albumin may

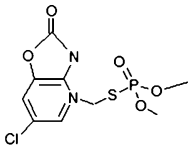
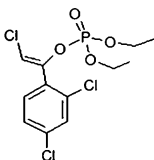
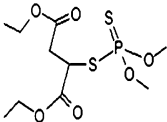
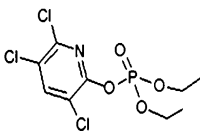
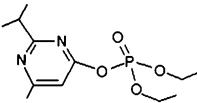
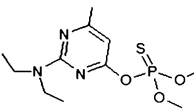
OP binding of albumin	
Low level	High level
Azamethiphos (oxon)	Chlorfenvinphos (oxon)
	
Malathion	Chlorpyrifos-oxon
	
	Diazinon-oxon
	
	Pirimiphos-methyl
	

Figure 6. Structures of low level and high level albumin-binding pesticides.

also be variable within the human population, and thereby influence an individual's ability to detoxify pesticides.

Once bound, an OP-adduct may undergo three fates: it may remain at the binding site until the protein is either turned over or actively degraded; spontaneous removal from the target by hydrolysis which results in the reformation of the native hydroxy amino acid; or ageing, whereby an alkyl side chain of the inhibitor is lost (dealkylation)

and the resulting enzyme-inhibitor complex becomes resistant to enzyme regeneration by fluoride ions or oximes. The relative rates of protein turnover/degradation, spontaneous reactivation, or ageing will determine the persistence of OP binding, and hence its usefulness as a biomarker of OP exposure.

The rates of ageing and spontaneous reactivation are specific to the bound phosphoryl group. For AChE, the stability and resistance to spontaneous reactivation of *O,O'*-dimethyl is less than *O,O'*-diethyl, which is less than *O,O'*-dipropyl bound counterparts, but structurally similar phosphoryl groups should all reactivate at approximately the same rate if bound to the same enzyme (Hobbiger 1951, Davison 1955). In the present study we examined dimethoxy- and diethoxy-albumin binding. The diethoxy pesticide binding was greater than that for the dimethoxy pesticide at 3 and 7 days after dosing which presumably represents enhanced resistance to spontaneous reactivation. However, for either pesticide the levels of pesticide binding were still significant 7 days after dosing, indicating good potential for both structures to provide post-exposure biomonitoring information. In the absence of sufficient data collection points it is difficult to accurately calculate the true half-lives of this pesticide to albumin binding, but since stable albumin adducts should follow a first-order rate of elimination (Granath et al. 1992, Troester et al. 2002), it is clear that pesticide albumin binding is not unstable but maintained at least as long as the approximately 3 day half-life of native rat albumin (Katz et al. 1961, Peters 1996).

By comparison, estimates of erythrocyte cholinesterase inhibited either *in vitro* or *in vivo* indicate a half-life of spontaneous reactivation of 1–2 h with dimethoxy OPs (Aldridge & Davison 1953, Vandekar & Heath 1956, Blaber & Creasey 1960, Worek et al. 1999), and approximately 2 days with diethoxy pesticides (Blaber & Creasey 1960, Mason et al. 2000). Other estimates of spontaneous reactivation of erythrocyte cholinesterase inhibited by dimethoxy pesticides have suggested a half-life of as much as 37 h (Mason et al. 2000). The levels of inhibitor used in these assays may contribute to the differences in the values attained for the dimethoxy phosphate esters due to sustained enzyme inhibition, and in practice, inhibition can be prolonged by ageing, but particularly for dimethoxy adducts plasma albumin clearly has a usefully long duration of binding relative to AChE. It will remain for future studies to evaluate the true rates of spontaneous reactivation of human albumin OP-bound *in vivo*.

Pesticide binding to albumin may not totally return to control levels due to ageing. However, the usefulness of albumin for retaining OP adduct information has been demonstrated from both *in vitro* and *in vivo* studies which have shown that albumin was resistant to ageing (and thereby susceptible to fluoride or oxime reactivation) after active-site occupancy by DFP, OPs or a number of nerve agents (Black et al. 1999, Adams et al. 2004, Li et al. 2007, Williams et al. 2007).

Human albumin has an estimated half-life of approximately 20 days (Peters 1996, Chaudhury et al. 2003), a life-span suitably persistent for biomarker sampling. However, at present we can only presume that the pesticide albumin binding stability that we observed for rats would also be seen in humans.

For comparison, the half-life of recovery of BuChE from the dimethoxy OP, dichlorvos, was approximately 12 days (Mason 2000), similar to the estimates of this protein's half-life (Munkner et al. 1961, Ostergaard et al. 1988). Erythrocyte AChE, undergoes a steady (linear) recovery from dichlorvos, returning to control levels after approximately 82 days – a little prior to the life-span of an erythrocyte (approximately

120 days), which demonstrates its value for monitoring relatively long-lasting OP inhibitions (Mason 2000).

In summary, the results described in this paper quantify binding of low levels of certain pesticides to rat serum albumin that persists beyond 7 days post-exposure, and suggest that the relatively high protein concentration of albumin, coupled to its relatively long half-life, make it a suitable biomonitor target for certain pesticides. Currently, use of gas chromatography to identify products from fluoride reactivation of BuChE-OP adducts provides a sensitive method for post-exposure OP detections (Polhuijs et al. 1997, van der Schans et al. 2004). However, this method is not without limitations, as OP-BuChE adducts are susceptible to ageing with associated loss of amenability to the reactivation process. Conversely, albumin adducted by OPs and some nerve agents is resistant to ageing thereby maintaining OP-adduct information which has been detected by a number of mass spectrometry-based methods (Black et al. 1999, Adams et al. 2004, Li et al. 2007, Williams et al. 2007). In future work it will be important to determine the *in vivo* threshold at which pesticide binding to albumin can be detected in order to further characterise albumin-binding sensitivity. However, mass spectrometric methods, or an enzyme-linked immunosorbent assay utilising antibodies specifically directed against albumin-OP adducts rather than the parent OP compound, will probably provide a more practicable means for quantifying the extent of OP-albumin binding, thereby avoiding the use of radioactive DFP.

The tendency of specific OP pesticides to differentially bind albumin and other non-AChE targets could be exploited via a panel of immune assays to gain information about the specific agents to which persons have been exposed. Thus malaoxon would adduct AChE but not albumin or acylpeptide hydrolase (Richards et al. 2000), whereas diazinon-oxon would bind albumin and AChE, but not acylpeptide hydrolase. This approach may alleviate the base-line problems associated with cholinesterase activity measurements that arise in the absence of pre-exposure values, and therefore help to provide a better means of correlating specific OP pesticide exposures with reported ill-health.

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